

Review article

Plant-derived natural compounds as promising anticancer agents in hematological malignancies

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Abstract

The latest treatments have improved outcomes for patients with hematological malignancies, but relapse, treatment resistance and particularly side effects still remain as common limitations of these treatments. Given the disadvantages of the existing conventional therapeutic methods, developing more effective drugs with less toxicity and side effects is of paramount importance. Medicinal herbs have historically proven their worth as a pool of potential therapeutic agents for leukemia and lymphoma, and today they still represent a rich source for the recognition of new drug leads. The role of the positive synergistic effects of plant-derived natural products and common chemotherapeutic drugs is also considered as one of the rational reasons for paying attention to the medicinal plants in recent chemoprevention and chemotherapeutic investigations. Noteworthy, targeted delivery of plant-derived natural products via the incorporation of nanoparticles or antibodies would be a major step to improve their bioavailability and then to increase their therapeutic effects. In this study, we reviewed plant-derived agents approved and/or under investigation for hematological malignancies.

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1. Introduction:

Leukemia and lymphoma are now curable in many cases due to the enormous improvements in the efficacy of chemotherapeutic drugs, however, significant adverse effects and chemoresistance have led to treatment failure and early relapse (1-4). Given the challenges of the existing conventional treatment

methods, developing novel and potent agents with less toxicity and side effects is one of the most important requirements in cancer treatment research. Despite the power of synthetic chemistry as an appealing field to discover and develop new therapeutic agents, the bioactive plant extracts

and their derivatives would be very worthwhile for detecting efficient and safe anti-tumor therapeutics (7, 8). American-Indians were the first to use extracts from the roots of mayapple, *Podophyllum peltatum*, as an effective treatment for skin cancers. The main constituent, podophyllotoxin, was the forerunner of the group of anticancer agents known as the podophyllins (9). Natural products have been used as traditional medicines throughout human history. Nowadays, about one-third of the common pharmaceuticals are natural products or their derivatives (10-12). Not only are a large number of chemotherapeutic agents derived from plants also many naturally occurring substances present in the plants have been identified as potential chemopreventive agents. Today, more researchers have focused on the utilization of phytochemicals such as curcumin, genistein, resveratrol, epigallocatechin-3-gallate, and sulforaphane for managing cancer (13-17). Additionally, the role of positive synergistic effects of secondary metabolites and chemo drugs should not be ignored as one of the rational reasons for paying attention to plant extracts in recent chemotherapeutic and chemoprevention investigations (18).

Despite the fact that natural products have been a rich source of compounds for drug discovery, their use has diminished in the past two decades in part because of technical barriers to screening them in high-throughput assays against molecular targets (19, 20). In some cancers, targeted therapies have greatly delayed tumor progression, and/or improved the life expectancy of patients. However, the vast majority of common cancers were found to be dependent not on a single “targetable” oncogenic activation. It is necessary to say that different types of leukemia and lymphoma are also shown to activate multiple adaptive mechanisms that either cause primary resistance to targeted drugs or help acquired resistance after only a few months of treatments (21). In spite of the predominant industrial focus on synthetic compound libraries, natural products still represent a valuable source for drug discovery given their described by Atanas G. Atanasov (22). Here, we review the history of and latest clinical findings on plant-derived agents (alone and/or in combination with other anticancer drugs) used or under investigation for patients with hematologic malignancies.

2. Anti-leukemic medicinal herbs and their derivatives

2.1. Chemopreventive agents

In general, cancer chemoprevention focuses on the identification of agents that specifically impact the early stages of cellular transformation (23, 24). Plant-derived phytochemicals have been found to have a wide range of anticancer activities in different stages of carcinogenesis, namely cancer initiation, proliferation, and progression (25). An extract derived from Leguminosae (*Cassia quinquangulata* Rich) was identified as a chemopreventive agent. Then, resveratrol was identified as its active component on the basis of bioassay-guided fractionation (15). In vitro and in vivo studies show that resveratrol by targeting cyclooxygenases (COXs) prevents carcinogenesis and reduce inflammation (15, 26-28). It has been reported that this phytochemical exhibited cytotoxicity in multiple myeloma cell lines through inhibition of NF- κ B, Akt, and STAT3 (29). Despite convincing pre-clinical evidence that resveratrol could aid in the treatment of patients with multiple myeloma, a clinical trial found that it caused several severe adverse events, the most prominent of which was renal failure (30). On the other hand, as resveratrol was proven to be safe in a phase II clinical trial, and it did not cause any nephrotoxicity in a phase I study for colorectal cancer patients, this adverse event seemed to be specific to multiple myeloma patients. A few clinical trials have shown that resveratrol has several targets within the cell such as caspase-3, and its efficacy is dependent on the type and stage of cancer, dosage levels, and treatment periods (31).

6-(methylsulfonyl) hexyl isothiocyanate (6-MITC) is another chemopreventive plant-derived product for leukemic that is derived from *Wasabia Japonica*. One study focusing on the evaluation of anti-leukemic potential of 6-MITC revealed its stronger cytotoxic effects on leukemic cells compared to healthy lymphocytes by inducing apoptosis, and blocking the cell cycle. Interestingly, this compound could induce differentiation of HL-60 cells into granulocytic and macrophage phenotypes (32). In general, despite the scientific importance and potential public health advantage, cancer chemoprevention has not been generally adopted in the clinic. In fact, the delay from the progression of cancer cell transformation to detectable cancer lesions may require years of patient data. However, utilizing malignant chemical biomarkers may illustrate a good way to recognize responses in patients taking phytochemicals.

2.2. Chemotherapeutic agents

2.2.1. Clinically approved agents

Of the 175 small molecules in the area of cancer, 131 (74.8%) are other than synthetic products with 85 (48.6%) actually being either natural products or their derived agents (22). Structures of antileukemic plant-derived agents and their sources were represented in Fig. 1. In this section, we will review the plant-derived chemotherapeutic agents which have been approved for therapeutic use for patients with hematological malignancies (Table 1).

Vinca alkaloids

Vinblastine and vincristine were first introduced in 1963 and 1965, respectively. They have contributed to long-term remissions and cures patients with childhood leukemia, Hodgkin's lymphoma and many other types of cancer (33, 34) (39). These compounds, known generally as vinca alkaloids, were isolated from the Madagascan periwinkle. Interactions of vinca alkaloids with tubulin, the major component of microtubules in the mitotic spindle, and the subsequent arrest of cells in mitosis (metaphase) are

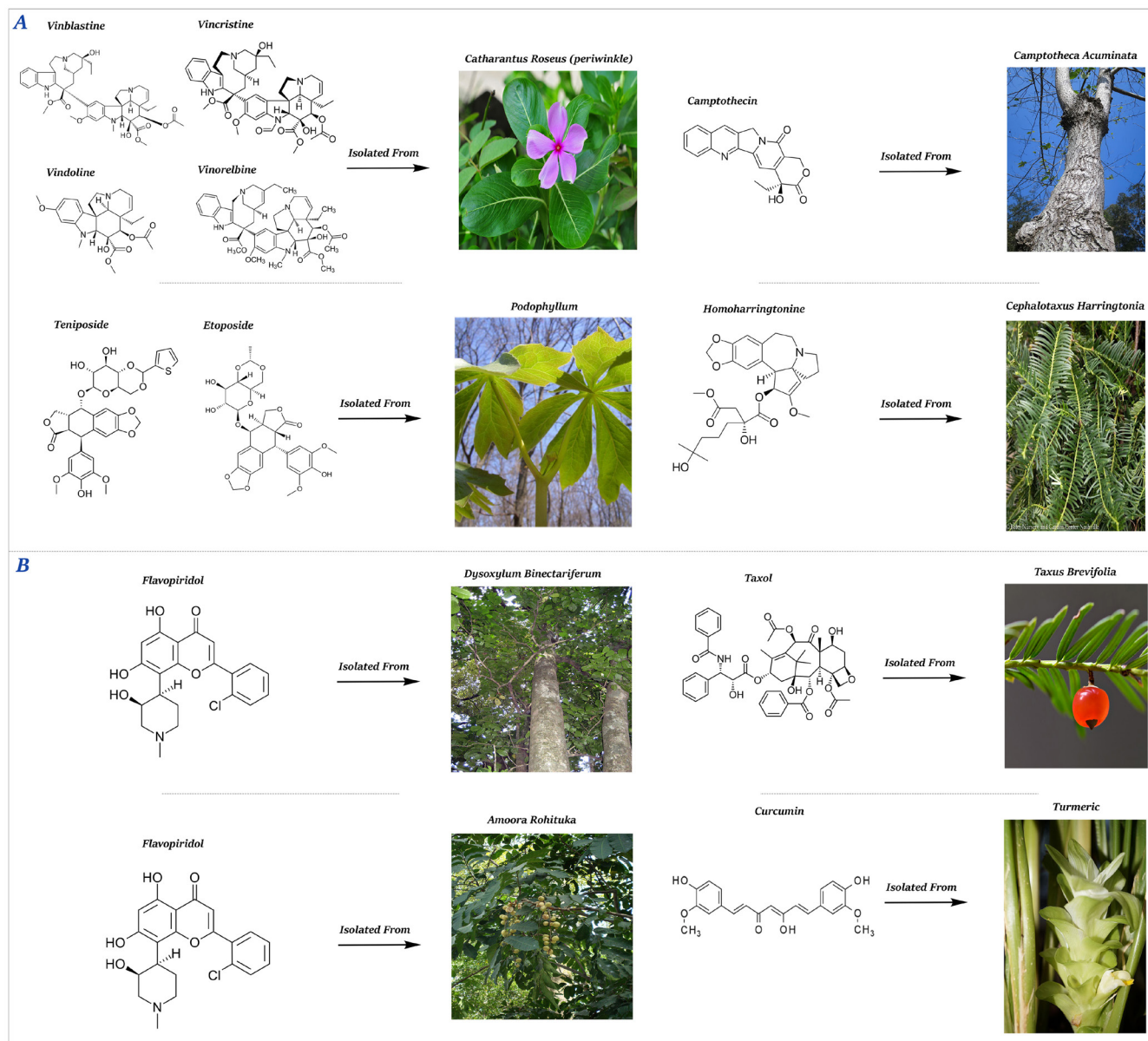


Figure 1: Structures of antileukemic plant-derived agents and their sources. A) Approved agents. B) Agents that are under clinical investigation.

Table 1: Plant-derived natural products approved for therapeutic use in leukemia and lymphoma.

Plant	Traditional use	Anticancer mechanism	Drug	Trade name	Approved by	Introduced	Hematologic malignancies
Catharanthus roseus	Anti-diabetic, anti-malaria and anti-tumor (65, 66)	Microtubule Inhibitor (Microtubule destabilizing)	Vincristine	Oncovin®	FDA	1963	ALL, CML, CLL, MM, HL, NHL
			Vinblastine	Alkaban-AQ®, Velban®	FDA	1965	HL, NHL, CML (blast crisis)
			Vindesine	Eldisine®	FDA	1979	Acute leukemia, lymphoma, HL
			Vinorelbine	Navelbine®	ARMC	1989	HL
			Liposomal vincristine	Marqibo®	FDA	2012	Ph- ALL
Podophyllum genus	Anthelmintic (67)	Inhibitor of topoisomerase II	Teniposide	Vumon®	FDA	1967	ALL, AML, CLL, MM, HL, NHL
			Etoposide	VP-16®	FDA	1980	Relapsed ALL in children
Camptotheca accuminata	Anti-tumor (67)	Inhibition of topoisomerase I	Topotecan	Hycampti®	ARMC	1996	AML, MDS, CML
Cephalotaxus harringtonia	Anthelmintic (67)	Inhibition of protein synthesis	Omacetaxine mepesuccinate (homoharringtonine)	Synribo®	FDA	2012	CML

Abbreviations: **ALL**, Acute Lymphoblastic Leukemia; **CML**, Chronic myeloid leukemia; **CLL**, Chronic lymphocytic leukemia; **MM**, Multiple myeloma; **HL**, Hodgkin lymphoma; **NHL**, Non-hodgkin lymphoma; **AML**, Acute myeloid leukemia; **Ph⁺ ALL**, Philadelphia chromosome negative acute lymphoblastic leukemia; **MDS**, Myelodysplastic syndrome.

generally accepted as key events in their mechanisms of action. Vinblastine sulfate is currently a component of a number of chemotherapy regimens for the treatment of Hodgkin's lymphoma and is also utilized in the therapy of bladder and breast cancers. Vincristine sulfate is used in various types of chemotherapy regimens for acute lymphoblastic leukemias (ALL) and lymphomas (33, 40). Selective chemical modifications of these two compounds have led to several structural analogues being approved in Europe for cancer treatment. Most notable of these analogous are vinorelbine (41) and vindesine (42, 43). These two compounds are semisynthetic derivatives of vinblastine with current or potential use in treating hematological malignancies (44). These agents are also used in combination with other cancer chemotherapeutic drugs for the treatment of a variety of cancers, including leukemia and lymphoma.

Podophyllotoxin derivatives

Epipodophyllotoxin is the naturally occurring epimer of podophyllotoxin (35) that was isolated as the active anti-tumor agent from the roots of various species of the genus *Podophyllum* (45). These medicinal herbs possess a long history of medicinal use by early Asian and American cultures, such as the treatment of skin cancers and warts (46). They inhibit topoisomerase II (topo II) and act by interfering with topoisomerase-mediated re-annealing of DNA (single- and double-strand) breaks, especially in the S and G2 phases of the cell cycle followed by accumulation of DNA damage and potent induction of caspase-dependent apoptosis (47). Epipodophyllotoxin is the parent of the two

clinically approved agents, etoposide and teniposide (35). Etoposide, etoposide phosphate (a water-soluble prodrug of etoposide), and Teniposide received Food and Drug Administration (FDA) approval in 1983, 1996, and 1993, respectively (48). Etoposide is in regular use for the effective treatment of testicular teratoma and small-cell lung cancer, whereas teniposide has efficacy against ALL and neuroblastoma in children, and non-Hodgkin's lymphoma and brain tumors in adults (9).

Camptothecin

Camptothecin, a cytotoxic pentacyclic alkaloid, was originally isolated from the bark and stem wood of *Camptotheca acuminata*, a tree indigenous to China in the early 1960s (49). This compound attracted immediate interest as a potential cancer chemotherapeutic agent due to its impressive activity against leukemia and various solid tumors in experimental systems (50). Due to negligible water solubility, this agent was advanced to clinical trials in water-soluble sodium salt form by National Cancer Institute (NCI) in the early 1970s, but it was dropped out of use in the late 1970s because of its lower efficacy and severe bladder- and myelo-toxicity.

Interest in camptothecin subsided for more than a decade (50). Revived attention resulted from the breakthrough discovery of DNA topoisomerase I (topo I) as a therapeutic target for camptothecin. This discovery put camptothecin back on the frontlines of anticancer drug development in the late 1980s (51-53). Not only camptothecin will help to understand the functions of the enzyme topo I for which it is a

specific inhibitor but also acts as a lead structure for the design of other molecules that selectively inhibit topoisomerases (54). Chemical manipulation of its structure subsequently produced analogues, including topotecan (Hycamtin) and irinotecan (Camptosar), that have been approved for use in the USA in 1996 (55). Topotecan, but not irinotecan, is used in clinical settings for a wide range of hematologic malignancies such as ALL, acute myeloblastic leukaemia (AML), myelodysplastic syndrome (MDS), and chronic myeloid leukemia (CML).

Homoharringtonine

Another plant-derived agent in clinical use is homoharringtonine that also called Omacetaxine mepesuccinate or Synribo. It is cephalotaxine ester isolated from *Cephalotaxus harringtonia* var. *drupacea* (Cephalotaxaceae), which is evergreen coniferous shrubs used in Chinese medicine for the treatment of cancer (56, 57). A racemic mixture of harringtonine and homoharringtonine has also been used successfully in China for the treatment of various types of leukemia. Clinically, purified homoharringtonine exhibits efficient inhibitory effects against AML (58, 59) and CML (60, 61) alone or combined with granulocyte colony-stimulating factor, cytarabine, or interferon- α . In addition, this agent has been reported to produce complete hematologic remission in patients with late chronic phase of CML. Homoharringtonine has been described as an inhibitor of protein synthesis; it inhibits chain elongation during translation by suppressing the substrate binding to the receptor site on the 60S ribosome subunit (62). Efferth et al found that homoharringtonine is more efficient in tumor cells with wild-type p53 in a high-throughput screening assay (with 55 NCI cell lines) (63). A recent study demonstrated that the possible mechanisms of homoharringtonine in myeloma may be the inhibition of Akt phosphorylation and Akt target genes including NF- κ B, X-linked inhibitor of apoptosis (XIAP), cellular inhibitor of apoptosis (cIAP), and Cyclin D1 (64). Inhibition of myeloid cell leukemia 1 (MCL1) protein synthesis, and induction of apoptosis were also reported in CML (61).

2.2.2. Agents under clinical investigation

Curcumin

Curcumin is the active compound in the traditional herbal medicine and dietary spice turmeric (*Curcuma*

longa) (68). This yellow spice has a long history of use in traditional remedies of China and India to treat common eye infections, and to treat bites, burns, acne and various skin diseases (69, 70). Curcumin has a surprisingly wide range of beneficial properties, including anti-inflammatory, antioxidant, and anti-cancerous activity (71). This ingredient is one of the most successful compounds investigated in recent years and is currently being assessed as both chemopreventive and chemotherapeutic agent. Curcumin is currently used in human clinical trials for a number of hematological malignancies such as multiple myeloma and MDS and also for a variety of disorders including colorectal cancer, pancreatic cancer, Alzheimer's disease, and psoriasis (25, 72-75). Several attempts combining curcumin with glucuronidation inhibitors such as piperine have also shown promising results to inhibit hepatic and intestinal metabolism (77). It inhibits cancer development and progression by targeting multiple signaling pathways including survival pathways such as those regulated by NF- κ B, Akt, and growth factors, extrinsic and intrinsic apoptosis pathways, and last but not least, metastatic and angiogenic pathways (68). Notably, curcumin may also function as a chemosensitizer and enhance the activity of other anticancer drugs, in part by disrupting pathways that lead to treatment resistance (76). It is remarkably non-toxic but exhibits limited bioavailability. Therefore, current research has shifted focus to improving bioavailability to overcome both the variability of absorption and rapid compound metabolism.

Taxol

Among the plant's natural products, Taxol is considered as the undoubted star, which shows efficacy against Kaposi's sarcoma and refractory breast and ovarian cancers. At present, Taxol is the best-selling anticancer agent that reached sales of US \$1.5 billion in 2000, which are still growing (78). The complex diterpene Taxol® (Paclitaxel) initially was isolated from the bark of *Taxus brevifolia* that historically had been used by several Native American tribes for the treatment of non-cancerous conditions. Leaves of *T. baccata* are also used in Ayurvedic (traditional Asiatic Indian) medicine system, with one reported application for the treatment of cancer (79, 80). Paclitaxel and also several key precursors (the baccatins) exist in the leaves of various *Taxus* species. Notably, the ready

semi-synthetic conversion of the relatively abundant baccatins to paclitaxel, as well as active paclitaxel analogs such as docetaxel, has provided a major, renewable natural source of this important class of drugs (81). Clinical trials of paclitaxel indicated the potential usefulness of this agent in the treatment of different types of hematological malignancies including ALL, AML, Hodgkin's and non-Hodgkin's lymphoma (Table 2). Unlike vinca alkaloids that destabilize microtubules, Taxol stabilizes them by binding to the beta-tubulin subunits during cell proliferation. Chromosomes are thus unable to achieve a metaphase spindle configuration. This blocks the progression of mitosis and prolonged activation of the mitotic checkpoint triggers apoptosis or reversion to the G₀-phase of the cell cycle without cell division. This allows the relationships between structure and activity to be established for hundreds of semi-synthetic analogues, with several more effective analogues being already in clinical trials. This natural product has therefore provided not only a potent drug but also the springboard for next developments (82-84).

Flavopiridol

Flavopiridol or alvocidib is being investigated for use in leukemia, lymphoma, lung, liver, and esophageal cancers. While flavopiridol is totally synthetic, the basis for its novel structure is the natural product rohitukine isolated from *Amoora rohituka* and *Dysoxylum binectariferum* (85-87). Flavopiridol is regarded as a pan-cyclin-dependent kinases (CDK) inhibitor that inhibits multiple CDKs by binding to the ATP-binding pocket of the kinase and then induces cell cycle arrest. Notably, flavopiridol appears to be the most potent agent against the CDK9 pathway which is dysregulated in AML (88). Initial clinical trials using flavopiridol in leukemia and lymphoma were disappointing (89, 90). Byrd and Grever found that the results could be explained by the large difference in protein binding of flavopiridol between fetal bovine serum (as typically used in cell culture) and human plasma. This observation and data from completed trials resulted in designing a schedule of flavopiridol administration with the goal of achieving brief high levels of free drug. Using this new schedule, this group showed that flavopiridol has potent activity in patients with drug-refractory chronic lymphocytic leukemia (CLL) (91). Now, flavopiridol is being evaluated in

a multi-center Phase II trial in relapsed/refractory CLL (92) and continues to be investigated in other hematologic malignancies such as AMLs, APL, CML, CLL, SLL, non-Hodgkin lymphoma, diffuse large B cell lymphoma, mantle cell lymphoma, etc. As a snapshot of the current state and perspective for future developments, an overview of advanced plant-derived small chemical entities that are used in clinical trials is presented in Table 2. Also, a schematic overview of the main targets of these agents was represented in Fig. 2.

3. Synergistic effects of plant-derived and chemotherapeutic agents

Combination therapy has gained momentum in oncology in recent years, with various studies demonstrating higher response rates with combination of drugs compared to monotherapies (113). In fact, the complex synergistic interactions between the drugs and herbs or natural products are supposed to be able to enhance therapeutic effects while reducing toxicity and drug resistance. The FDA recently approved therapeutic combination regimens that illustrated superior efficacy and safety to monotherapies. In addition, the National Cancer Institute has recently highlighted combination therapy as a top research priority in oncology. The major benefit of combination therapies is that they reduce the development of drug resistance since cancer is less likely to have resistance to multiple drugs simultaneously. This approach may seem costlier than monotherapy in the short term, but when it is used appropriately, it causes significant savings: lower treatment failure rate, lower case-fatality ratios, slower development of resistance, and fewer adverse effects than monotherapy (114). Since patients treated with doxorubicin that has potent antineoplastic activity over acute leukemia suffer from acute and delayed side effects (115) and also demonstrate acquired drug resistance, a recent study investigated the effect of the combination of doxorubicin and several plants' secondary metabolites (sanguinarine, epigallocatechin-3-gallate, etc.) that modulate multi drug resistance (MDR) on drug-resistant leukemia cells. Notably, combination of nontoxic concentrations of individual natural compound with doxorubicin was shown to significantly sensitize leukemic cells, and significantly enhance the cytotoxicity of doxorubicin (116). Some examples of natural products used in combination with chemotherapeutic drugs that are in phase I-IV clinical trials are summarized in Table 3.

Table 2. Plant-derived natural products used in clinical trials for leukemia and lymphoma.

Plant	Traditional use	Lead component(s)	Drug	Other name	Mechanism of action	Hematologic neoplasia	Clinical trial
Curcuma longa	Urinary diseases, diseases of liver, jaundice and cancer (93, 94)	Curcumin	Curcumin	Turmeric Yellow	Anti-inflammatory, anti-oxidative	Maintenance therapy in MM MM T-cell Lymphoma Patients	Phase II Phase II Phase II
Taxus brevifolia	Some non-cancerous conditions and cancer (95)	Paclitaxel	Paclitaxel	Taxol	Microtubule inhibition	Refractory acute leukemia, CML Recurrent and refractory lymphoma NHL	Phase II Phase I/II Phase II
			Ortataxel	IDN-5109, BAY-59-8862	Microtubule inhibition		
Red grapes	Hyperlipidemia, fatty liver, diabetes, atherosclerosis and aging (96-100)	Resveratrol	Resveratrol	SRT501	Cell cycle arrest and apoptosis	MM	Phase II
Tripterygium wilfordii	Arthritis, muscle and skeletal injury, and skin diseases (101)	Triptolide	Minnelide™ 001	Triptolide O-Methyl	Apoptosis inducing by HSP70	AML	Phase I
Camellia sinensis (Green and white tea)	Flatulence, fever, and diabetes (102, 103)	Catechin	Catechin	Cianidanol	Antioxidant	MM and plasma cell neoplasm	Phase II
Pomegranate	Microbial infection, inflammation, diabetes and cancer (104-106)	Ellagic acid	Ellagic acid	Benzoaric acid	Anti-proliferative, antioxidant	MDS	Phase II
Maclura pomifera	Disease of eye(107)	Morin	Morin	Aurantica	Antiproliferative	AML	Phase I
Amoora rohituka	Microbial infection and diseases of liver (108)	Rohitukine	Alvocidib	Flavopiridol, DSP-2033	CDK9 inhibition	AML CML or Lymphocytic Lymphoma CLL or APL CLL or SLL AML, ALL, or CML CLL or APL Arising From CLL CLL Lymphoma or MM MCL or DLBCL Lymphomas Intermediate-Grade or High-Grade NHL or MCL	Phase I Phase II Phase II Phase I Phase I Phase II Phase II Phase II Phase II Phase I Phase II
			Voruciclib	P1446A	CDK4 and CDK6 inhibition	B-cell malignancy CLL	Phase I Phase I

Abbreviations: **MM**, Multiple myeloma; **CML**, Chronic myeloid leukemia; **NHL**, Non-hodgkin lymphoma; **AML**, Acute myeloid leukemia; **MDS**, Myelodysplastic syndrome; **CLL**, Chronic lymphocytic leukemia; **APL**, Acute promyelocytic leukemia; **SLL**, Small lymphocytic lymphoma; **MCL**, Mantle cell lymphoma; **DLBCL**, Diffuse large B cell lymphoma.

4. Targeted drug delivery system of medicinal herbs

Targeted drug delivery system refers to a method using different carriers to change the effective components extracted from natural products into agents which can directly concentrate on the target site. This system is an ideal delivery approach and it can reduce the adverse reactions and improve the pharmacological effects [1]. There are different carriers which two of the most important ones used for hematological malignancies are discussed in this review.

4.1. Nanoparticle delivery

The biggest issue with the use of natural products in disease treatment that makes researchers interested in targeted therapy with nanoparticle is their low bioavailability, which has caused problems in clinical trials. Highly lipophilic compounds such as curcumin, resveratrol, and epigallocatechin-3-gallate (EGCG) are not ideal for drug delivery because they do not dissolve well in the bloodstream. These compounds have low bioavailability, and therefore, large quantities of the compounds must be administered in order to achieve

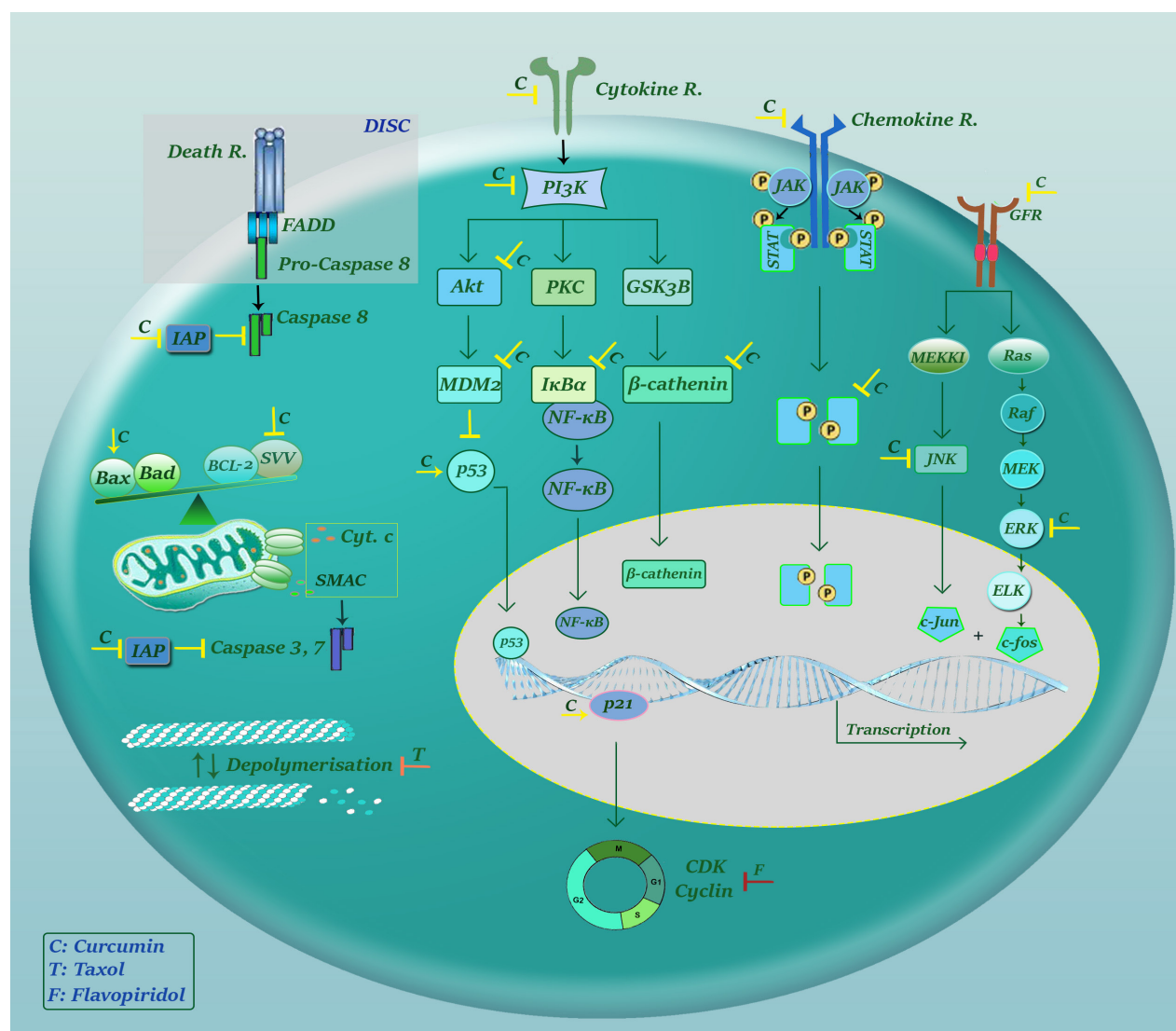


Figure 2: Schematic overview of the main targets of plant-derived natural compounds that are under investigation. Three compounds are shown by C (Curcumin), T (Taxol) and F (Flavopiridol). Curcumin is able to target several mechanisms that are involved in hematological malignancies. It targets several main signaling pathways including Ras/Raf/MEK/ERK pathway, PI3K/Akt pathway, extrinsic and intrinsic apoptosis pathways and JAK/STAT pathway. Taxol stabilizes the microtubule polymer and protects it from disassembly, which in turn blocks the progression of mitosis. Flavopiridol inhibits different cyclins and cyclin-dependent kinases (CDK) and then induces cell cycle arrest and apoptosis. GFR: Growth factor receptor; MEK1: Mitogen-activated protein kinase kinase 1; JNK: c-Jun N-terminal kinase; JAK: Janus kinase; STAT: Signal transducer and activator of transcription; PI3K: Phosphatidylinositol 3-kinase; GSK3B: Glycogen synthase kinase 3 beta; PKC: Protein kinase C; NF-κB: Nuclear factor-κB; MDM2: Mouse double minute 2 homolog; DISC: Death-inducing signaling complex; FADD: Fas-associated protein with death domain; IAP: Inhibitor of apoptosis; Bcl-2: B-cell lymphoma 2; Bax: Bcl-2-associated X protein; Bad: Bcl-2-associated agonist of cell death; SVV: Survivin; Cyto. C: Cytochrome c; SMAC: Second mitochondria-derived activator of caspases; CDK: Cyclin-dependent kinase.

the optimum dose in the body. Nanotechnology brings multiple advantages to the targeted delivery of natural products in the treatment of cancers. The utilization of nanoparticles along with natural components can increase the bioavailability, targeting, and controlled-release profiles of these agents (10). One of the first studies that employed nanoparticles in the field of delivery of leukemia and lymphoma drugs was done in the 1990s (117, 118). Preliminary studies were concerned with pharmacokinetic behavior and plasma concentration of a vincristine sulfate liposome system.

that was FDA-approved under the name Marqibo® as an injected nano-drug used against relapsed Ph-ALL (Philadelphia chromosome-negative ALL) in adults (119). Recently, the safety profile and clinical benefit of Marqibo® was assessed for the adolescent young adult population and for children, and it was reported to exhibit safety and efficiency profiles similar to those obtained for adult patients (120, 121). Another study that demonstrated the applicability of nanoparticles for ALL was done by Changhui Liu et al (122). The targeted nano-containers were able to deliver and

Table 3. Plant-derived natural products used in combination with other cytotoxic agents in clinical trials for leukemia and lymphoma.

Plant	Drug	Other name	Mechanism of action	In combination with	Hematologic Neoplasia	Clinical trial
Taxus brevifolia	Paclitaxel	Taxol	Microtubule Inhibition	Carboxyamidotriazole Topotecan and Filgrastim-SD/01 Avelumab, bevacizumab, capecitabine, cyclophosphamide, 5-Fluorouracil (5-FU), leucovorin, nab-paclitaxel, Lovaza, Oxaliplatin, Rituximab, ALT-803, ETBX-061 and haNK Monoclonal antibody therapy and Cyclosporine Estramustine L-778,123 Ifosfamide and Teniposide Cyclophosphamide and Dexamethasone Lenalidomide (Revlimid)	Refractory lymphomas Relapsed and refractory aggressive NHL Relapsed CD20-positive NHL Recurrent or refractory NHL Relapsed or refractory NHL Recurrent or refractory lymphomas Relapsed NHL Relapsed or refractory MM Relapsed/refractory MM	Phase II Phase II Phase II Phase I Phase I Phase I Phase II Phase II Phase II
Amoora rohituka	Alvocidib	Flavopiridol, DSP-2033	CDK9 inhibition	Venetoclax Cytarabine/Daunorubicin Cytarabine, and Mitoxantrone Hydrochloride or Cytarabine and Daunorubicin Hydrochloride Cytarabine and Mitoxantrone Cytarabine, and Mitoxantrone Vorinostat Fludarabine Phosphate, and Rituximab	AML AML AML Newly diagnosed acute leukemia Acute Leukemia Relapsed/refractory acute leukemia or CML LPDs or MCL	Phase I Phase I Phase II Phase II Phase I Phase I
Maytenus serrata	Maytansine	Maitansine	Microtubule assembly inhibition	SAR3419 Lorvotuzumab Mertansine (IMGN901) LOP628	Leukemia ALL AML	Phase II Phase II Phase I
Pomegranate	Ellagic acid	Benzoic acid	Anti-proliferative, apoptotic, antioxidant	Dietary intervention: 33 Omega 3 fatty acid, L-Selenomethionine, Allicin, resveratrol, quercetin, Epigallocatechin gallate	Follicular lymphoma	Phase II
Salvia miltiorrhiza	Tanshinone I	TTE-50	Anti-proliferative and apoptotic	ATO, RIF, ATRA, mitoxantrone, Ara-C, MTX and 6MP	APL	Phase IV

Abbreviations: **NHL**, Non-hodgkin lymphoma; **MM**, Multiple myeloma; **AML**, Acute myeloid leukemia; **LPDs**, Lymphoproliferative disorders; **CML**, Chronic myeloid leukemia; **MCL**, Mantle cell lymphoma; **ALL**, Acute lymphoblastic leukemia; **APL**, Acute promyelocytic leukemia.

selectively release the entrapped paclitaxel drug into ALL cells (CEM and Ramos). Not only this drug delivery system targets ALL cells and leaves normal cells but also decreases the occurrence of some side effects of paclitaxel, leading toward better therapeutic efficacy in CEM-tumor-bearing mice (123). In addition, solid-lipid (one type of nanoparticle) formulation of a natural chemotherapeutic agent, AP9-cd, which was isolated from a dry waste wood powder of Cedrus deodara, was recently reported (124). This formulation showed improved anti-leukemic effect both in vitro and in vivo in comparison with conventional drug systems in drug efficacy, pharmacokinetics, and biodistribution aspects (123).

Although there are many advantages to apply nanotechnology for better delivery of natural components, it is not without challenges. Nanoparticles' potential toxicity is a major concern, partly because they can cross biological membranes, such as cellular membranes (126, 127). Another challenge that arises when using nanoparticles as drug delivery agents

is that the nanoparticle will undergo changes in the body. This must be taken into account because these changes could affect the bioavailability, targeting, and release kinetics of the drug (10). Nonetheless, many researchers believe that nanomedicines will shift the paradigm of cancer treatment and that the true goal of cancer nano-medicine will become a reality in the foreseeable future.

4.2. Antibody-drug conjugates (ADCs)

A burgeoning class of monoclonal antibody therapies, called antibody-drug conjugates (ADCs), delivers dual therapies in a single cancer cell-killing package (128). The localized delivery and release of the cytotoxic agent attached to a monoclonal antibody within or near malignant cells allow for the targeted delivery of a potent cytotoxic agent to antigen-positive cancer cells while reducing damage to antigen-negative normal cells (129). Mylotarg (gemtuzumab ozogamicin; GO) is a conjugate of a humanized monoclonal antibody that targets the CD33 antigen on leukemia cells with

a prodrug form of calicheamicin, a natural product derived from the bacterium *Micromonospora echinospora* (a species of bacteria) (130, 131). GO was the first antibody-drug conjugate approved for the treatment of AML by the FDA via the accelerated approval process in 2000 (132,133). Once bound to the leukemia cells, the conjugate is internalized and then breaks down, allowing calicheamicin to target a specific region of the minor groove of DNA (134). Two other immunoconjugates brentuximab vedotin (Adcetris®) (135) and trastuzumab emtansine (Kadcyla®) (136) were marketed in 2011 and 2013, respectively. Adcetris® targets the protein CD30, which is expressed in systemic anaplastic large cell lymphoma and classical Hodgkin lymphoma. This antibody is conjugated to a fully synthetic analogue of the antimitotic agent dolastatin, a linear pentapeptide originally isolated from the extracts of the sea hare *Dolabella auriculari* (a species of sea slug) (137).

Despite the importance of antibody-conjugated natural products derived from different sources (bacteria, sea slug, etc.) in the treatment of hematological malignancies, there has been no approved conjugated drug that derived from plant sources. Nevertheless, the promising news in this era is that many studies are now investigating conjugated plant-derived natural products in pre-clinical levels. For example, a study was designed to determine whether antibodies against human chorionic gonadotropin beta (hCG) conjugated to curcumin can selectively kill leukemic cells expressing hCG. This study was carried out on MOLT-4 and U-937 cells expressing hCG and on peripheral blood leukocytes of AML patients. The resulting data showed that antibody against hCG linked to curcumin has potential for therapy of hCG-expressing leukemia (138). Taken together, application of ADC strategy is a renewed interest in plant-derived natural products that are highly cytotoxic agents but suffer from lack of good drug-like properties and narrow safety margins when systemically administered as common chemotherapeutic drugs.

5. Conclusion

Since there are some important challenges related to using conventional therapeutic drugs for patients with leukemia and lymphoma, it is considered so necessary to find new agents that would be more efficient.

Medicinal plants have historically been a rich source for successful anti-leukemic drugs, and still represent a rich pool for the identification of new pharmacological leads. We are accepting that nature has already carried out the combinatorial chemistry; all we have to do is sometimes refine the structures. In addition, since the efficient in vivo uptake of leukemic cells requires better targeting moieties to improve cellular uptake and reduced toxicity to contiguous normal cells, many researchers are now investigating to find more targeted agents. Targeted therapy in leukemia and lymphoma treatment has made immense progress towards generating molecules that show great clinical benefit, resulting in FDA approval of some drugs such as Marqibo and the clinical development of many more. Clearly, there is ample scope for new discoveries to be made of natural products from plants with promising activities against leukemia and lymphoma. Finally, since most of today's natural products in clinical use were discovered through a routine examination of plants, so serendipity is still an important route of discovery.

Conflict of interest

The authors declare no conflict of interest.

Research involving human participants and/or animals

This article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent

For this type of study, informed consent is not required.

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References

1. Van Vlierberghe P, Ferrando AJTJoci. The molecular basis of T cell acute lymphoblastic leukemia. 2012;122(10):3398-406.
2. Uchida J, Okada H, Ohguchi N, Kawa G, Koyama Y, Mikami O, et al. Comparison of side effects caused by intra-arterial and intravenous infusion of M-VAC (methotrexate, vinblastine, adriamycin and cisplatin) for urothelial cancer. 1997;43(9):637-40.
3. Walker FE, editor Paclitaxel (TAXOL®): side effects and patient education issues. Seminars in oncology nursing; 1993: Elsevier.
4. Brockmann B, Geschke E, Schmidt U, Ebeling KJGuF. Therapeutic results and toxic side effects of the cytostasan, adriamycin and vincristine combination as second line therapy in metastatic breast cancer. 1991;51(5):383-6.

5. M Lucas D, C Still P, Bueno Perez L, R Grever M, Douglas Kinghorn AJCdt. Potential of plant-derived natural products in the treatment of leukemia and lymphoma. 2010;11(7):812-22.
6. Elnakady YA, Rushdi AI, Franke R, Abutaha N, Ebaid H, Baabbad M, et al. Characteristics, chemical compositions and biological activities of propolis from Al-Bahah, Saudi Arabia. 2017;7:41453.
7. Kaur M, Singh RP, Gu M, Agarwal R, Agarwal CJCCR. Grape seed extract inhibits in vitro and in vivo growth of human colorectal carcinoma cells. 2006;12(20):6194-202.
8. Widodo N, Kaur K, Shrestha BG, Takagi Y, Ishii T, Wadhwa R, et al. Selective killing of cancer cells by leaf extract of Ashwagandha: identification of a tumor-inhibitory factor and the first molecular insights to its effect. 2007;13(7):2298-306.
9. Mann JJNRC. Natural products in cancer chemotherapy: past, present and future. 2002;2(2):143.
10. Watkins R, Wu L, Zhang C, Davis RM, Xu BJljon. Natural product-based nanomedicine: recent advances and issues. 2015;10:6055.
11. Manly SP, Padmanabha R, Lowe SE. Natural products or not? How to screen for natural products in the emerging HTS paradigm. High Throughput Screening: Springer; 2002. p. 153-68.
12. Strohl WRJDDt. The role of natural products in a modern drug discovery program. 2000;5(2):39-41.
13. Surh Y-JJNRC. Cancer chemoprevention with dietary phytochemicals. 2003;3(10):768.
14. Lamartiniere CA, Moore J, Holland M, Barnes SJPotSfEB, Medicine. Neonatal genistein chemoprevents mammary cancer. 1995;208(1):120-3.
15. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. 1997;275(5297):218-20.
16. Paschka AG, Butler R, Young CY-FJCl. Induction of apoptosis in prostate cancer cell lines by the green tea component, (-)-epigallocatechin-3-gallate. 1998;130(1-2):1-7.
17. Li Y, Zhang T, Korkaya H, Liu S, Lee H-F, Newman B, et al. Sulforaphane, a dietary component of broccoli/broccoli sprouts, inhibits breast cancer stem cells. 2010;1078-0432. CCR-09-2937.
18. Bahmani F, Esmaili S, Bashash D, Dehghan-Nayeri N, Mashati P, Gharehbaghian AJB, et al. Centaurea albonitens extract enhances the therapeutic effects of Vincristine in leukemic cells by inducing apoptosis. 2018;99:598-607.
19. Harvey AL, Edrada-Ebel R, Quinn RJJNRDD. The re-emergence of natural products for drug discovery in the genomics era. 2015;14(2):111.
20. Atanasov AG, Waltenberger B, Pferschy-Wenzig E-M, Linder T, Wawrosch C, Uhrin P, et al. Discovery and resupply of pharmacologically active plant-derived natural products: a review. 2015;33(8):1582-614.
21. Basmadjian C, Zhao Q, Bentouhami E, Djehal A, Nebigil CG, Johnson RA, et al. Cancer wars: natural products strike back. 2014;2:20.
22. Newman DJ, Cragg GMJJJonp. Natural products as sources of new drugs over the 30 years from 1981 to 2010. 2012;75(3):311-35.
23. Sapienza C, Issa J-PJAr. Diet, nutrition, and cancer epigenetics. 2016;36:665-81.
24. Collins AR, Azqueta A, Langie SAJEjon. Effects of micronutrients on DNA repair. 2012;51(3):261-79.
25. Kotecha R, Takami A, Espinoza JLJO. Dietary phytochemicals and cancer chemoprevention: a review of the clinical evidence. 2016;7(32):52517.
26. Alayev A, Berger SM, Holz MKJAotNYAoS. Resveratrol as a novel treatment for diseases with mTOR pathway hyperactivation. 2015;1348(1):116-23.
27. Kulkarni SS, Cantó CJBeBA-MBoD. The molecular targets of resveratrol. 2015;1852(6):1114-23.
28. Pezzuto JMJPB. Resveratrol as an inhibitor of carcinogenesis. 2008;46(7-8):443-573.
29. Jazirehi AR, Bonavida BJMct. Resveratrol modifies the expression of apoptotic regulatory proteins and sensitizes non-Hodgkin's lymphoma and multiple myeloma cell lines to paclitaxel-induced apoptosis. 2004;3(1):71-84.
30. Popat R, Plesner T, Davies F, Cook G, Cook M, Elliott P, et al. A phase 2 study of SRT 501 (resveratrol) with bortezomib for patients with relapsed and or refractory multiple myeloma. 2013;160(5):714-7.
31. Berman AY, Motechin RA, Wiesenfeld MY, Holz MKJNpo. The therapeutic potential of resveratrol: a review of clinical trials. 2017;1(1):35.
32. Lenzi S, Cocchi V, Malaguti M, Barbalace MC, Marchionni S, Hrelia S, et al. 6-(Methylsulfonyl) hexyl isothiocyanate as potential chemopreventive agent: molecular and cellular profile in leukaemia cell lines. 2017;8(67):111697.
33. Guéritte F, Fahy J. The vinca alkaloids. Anticancer agents from natural products: CRC Press; 2005. p. 131-43.
34. Cimino G, De Rosa S, De Stefano SJE. Antiviral agents from a gorgonian, Eunicella cavolini. 1984;40(4):339-40.
35. Newman DJ, Cragg GM, Snader KMJNpr. The influence of natural products upon drug discovery. 2000;17(3):215-34.
36. Pezzuto JMJBp. Plant-derived anticancer agents. 1997;53(2):121-33.
37. Beer CT, Cutts JH, Noble RL. Vincalukoblastine. Google Patents; 1963.
38. Cutts J, Beer C, Noble RJCR. Biological properties of Vincalukoblastine, an alkaloid in Vinca rosea Linn, with reference to its antitumor action. 1960;20(7):1023-31.
39. Karon M, Freireich EJ, Frei E, Taylor R, Wolman IJ, Djerassi I, et al. The role of vincristine in the treatment of childhood acute leukemia. 1966;7(3):332-9.
40. Catterall W, Goodman MK. Gildmans. The pharmacological basis of therapeutics 11 th edition. Chapter 14. New York: Mc. Graw Hill; 2006.
41. Darke P, Leu C, Davis L, Heimbach J, Diehl R, Hill W, et al. Human immunodeficiency virus protease. Bacterial expression and characterization of the purified aspartic protease. 1989;264(4):2307-12.
42. Wiley RA, Rich DHJMrr. Peptidomimetics derived from natural products. 1993;13(3):327-84.
43. Fahy JJCpd. Modifications in the upper or velbenamine part of the vinca alkaloids have major implications for tubulin interacting activities. 2001;7(13):1181-97.
44. Williams DH, Stone MJ, Hauck PR, Rahman SKJJoNP. Why are secondary metabolites (natural products) biosynthesized? 1989;52(6):1189-208.
45. Canel C, Moraes RM, Dayan FE, Ferreira DJP. Podophyllotoxin. 2000;54(2):115-20.
46. Snader M, McCloud TGJESND. Ethnobotany and drug discovery: the experience of the US National Cancer Institute. 1994;185:178.
47. Van Maanen J, Retel J, De Vries J, Pinedo HJJJotNCI. Mechanism of action of antitumor drug etoposide: a review. 1988;80(19):1526-33.

48. Hande KJEjoc. Etoposide: four decades of development of a topoisomerase II inhibitor. 1998;34(10):1514-21.
49. Wall ME, Wani MC, Cook C, Palmer KH, McPhail Aa, Sim GJJotACS. Plant antitumor agents. I. The isolation and structure of camptothecin, a novel alkaloidal leukemia and tumor inhibitor from camptotheca acuminata1, 2. 1966;88(16):3888-90.
50. Liu YQ, Li WQ, Morris-Natschke SL, Qian K, Yang L, Zhu GX, et al. Perspectives on biologically active camptothecin derivatives. 2015;35(4):753-89.
51. Potmesil M, Kohn KW. DNA topoisomerases in cancer: Oxford University Press, USA; 1991.
52. Potmesil M, Giovanella B. Preclinical development of 20 (S)-camptothecin, 9-aminocamptothecin, and other analogs Potmesil M. Pinedo H. eds.. Camptothecins: New Anticancer Agents: 51-57. CRC Press Boca Raton, FL; 1995.
53. Hsiang Y-H, Hertzberg R, Hecht S, Liu LJJoBC. Camptothecin induces protein-linked DNA breaks via mammalian DNA topoisomerase I. 1985;260(27):14873-8.
54. Chen AY, Liu LFJArop, toxicology. DNA topoisomerases: essential enzymes and lethal targets. 1994;34(1):191-218.
55. Jonsson E, Dhar S, Jonsson B, Nygren P, Graf W, Larsson RJEJoC. Differential activity of topotecan, irinotecan and SN-38 in fresh human tumour cells but not in cell lines. 2000;36(16):2120-7.
56. Itokawa H, Wang X, Lee K-H. Homoharringtonine and related compounds. Anticancer agents from natural products: CRC Press; 2005. p. 56-79.
57. Efferth T, Li PC, Konkimalla VSB, Kaina BJTimm. From traditional Chinese medicine to rational cancer therapy. 2007;13(8):353-61.
58. Feldman E, Seiter K, Ahmed T, Baskind P, Arlin ZJL. Homoharringtonine in patients with myelodysplastic syndrome (MDS) and MDS evolving to acute myeloid leukemia. 1996;10(1):40-2.
59. Kantarjian HM, Keating MJ, Walters RS, Koller CA, McCredie KB, Freireich EJJ. Phase II study of low-dose continuous infusion homoharringtonine in refractory acute myelogenous leukemia. 1989;63(5):813-7.
60. Quintás-Cardama A, Kantarjian H, Garcia-Manero G, O'Brien S, Faderl S, Estrov Z, et al. Phase I/II study of subcutaneous homoharringtonine in patients with chronic myeloid leukemia who have failed prior therapy. 2007;109(2):248-55.
61. Chen R, Guo L, Chen Y, Jiang Y, Wierda WG, Plunkett WJB. Homoharringtonine reduced Mcl-1 expression and induced apoptosis in chronic lymphocytic leukemia. 2011;117(1):156-64.
62. FRESNO M, JIMÉNEZ A, VÁZQUEZ DJEjob. Inhibition of translation in eukaryotic systems by harringtonine. 1977;72(2):323-30.
63. Efferth T, Sauerbrey A, Halatsch M-E, Ross DD, Gebhart EJJN-Ssaop. Molecular modes of action of cephalotaxine and homoharringtonine from the coniferous tree Cephalotaxus hainanensis in human tumor cell lines. 2003;367(1):56-67.
64. Meng H, Yang C, Jin J, Zhou Y, Qian WJL, lymphoma. Homoharringtonine inhibits the AKT pathway and induces in vitro and in vivo cytotoxicity in human multiple myeloma cells. 2008;49(10):1954-62.
65. Cragg GM, Newman DJ. Plants as a source of anti-cancer agents. Journal of ethnopharmacology. 2005;100(1):72-9.
66. Botsaris ASJJoe, Ethnomedicine. Plants used traditionally to treat malaria in Brazil: the archives of Flora Medicinal. 2007;3(1):18.
67. Efferth T, Li PC, Konkimalla VSB, Kaina B. From traditional Chinese medicine to rational cancer therapy. Trends in molecular medicine. 2007;13(8):353-61.
68. Hatcher H, Planalp R, Cho J, Torti F, Torti SJC, sciences ml. Curcumin: from ancient medicine to current clinical trials. 2008;65(11):1631-52.
69. Thakur R, Puri HS, Husain AJLCIoM, Aromatic Plants 585p.-illus. ciEIG. Major medicinal plants of India. 1989;6.
70. Ammon HP, Wahl MAJPM. Pharmacology of Curcuma longa. 1991;57(01):1-7.
71. Rainey N, Motte L, Aggarwal B, Petit P. Curcumin hormesis mediates a cross-talk between autophagy and cell death. Nature Publishing Group; 2015.
72. Yang C, Su X, Liu A, Zhang L, Yu A, Xi Y, et al. Advances in clinical study of curcumin. 2013;19(11):1966-73.
73. Epelbaum R, Schaffer M, Vziel B, Badmaev V, Bar-Sela GJN, cancer. Curcumin and gemcitabine in patients with advanced pancreatic cancer. 2010;62(8):1137-41.
74. Dhillon N, Aggarwal BB, Newman RA, Wolff RA, Kunnumakkara AB, Abbruzzese JL, et al. Phase II trial of curcumin in patients with advanced pancreatic cancer. 2008;14(14):4491-9.
75. Sharma RA, McLelland HR, Hill KA, Ireson CR, Euden SA, Manson MM, et al. Pharmacodynamic and pharmacokinetic study of oral Curcuma extract in patients with colorectal cancer. 2001;7(7):1894-900.
76. Garg AK, Buchholz TA, Aggarwal BBJA, signaling r. Chemosensitization and radiosensitization of tumors by plant polyphenols. 2005;7(11-12):1630-47.
77. Shoba₁ G, Joy₁ D, Joseph₁ T, Rajendran₂ MMR, Srinivas₂ PJPm. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. 1998;64:353-6.
78. Goodman J, Walsh V. The story of taxol: nature and politics in the pursuit of an anti-cancer drug: Cambridge University Press; 2001.
79. Hartwell LJL. Plants used against cancer. A survey. [Continued.]. 1970;33:97-194.
80. Kapoor LJC, USA. CRC handbook of Ayurvedic plants. 1990;183.
81. Cortes JE, Pazdur RJJoCO. Docetaxel. 1995;13(10):2643-55.
82. Schiff PB, Fant J, Horwitz SBJN. Promotion of microtubule assembly in vitro by taxol. 1979;277(5698):665.
83. Chesnoff SJN. The use of Taxol as a trademark. 1995;374(6519):208-.
84. Kingston DGJCC. Taxol, a molecule for all seasons. 2001(10):867-80.
85. Endo AJTjoA. Monacolin K, a new hypocholesterolemic agent that specifically inhibits 3-hydroxy-3-methylglutaryl coenzyme A reductase. 1980;33(3):334-6.
86. Harmon AD, Weiss U, Silvertown JJTL. The structure of rohitukine, the main alkaloid of Amoorarohituka (Syn. Aphanamixispolystachya)(meliaceae). 1979;20(8):721-4.
87. Newman D. Anticancer Agents from Natural Sources. Cragg GM, Kingston DGI and Newman DJ. CRC Press/Taylor & Francis, Boca Raton, FL, pp553-571; 2005.
88. de Azevedo WF, Canduri F, da Silveira NJFJB, communications br. Structural basis for inhibition of cyclin-dependent kinase 9 by flavopiridol. 2002;293(1):566-71.
89. Flinn IW, Byrd JC, Bartlett N, Kipps T, Gribben J, Thomas D, et al. Flavopiridol administered as a 24-hour continuous infusion in chronic lymphocytic leukemia lacks clinical activity. 2005;29(11):1253-7.
90. Byrd JC, Peterson BL, Gabrilove J, Odenike OM, Grever MR, Rai K, et al. Treatment of relapsed chronic lymphocytic leukemia by 72-hour continuous infusion or 1-hour bolus infusion of flavopiridol: results from Cancer and Leukemia Group B study 19805. 2005;11(11):4176-81.

91. Byrd JC, Lin TS, Dalton JT, Wu D, Phelps MA, Fischer B, et al. Flavopiridol administered using a pharmacologically derived schedule is associated with marked clinical efficacy in refractory, genetically high-risk chronic lymphocytic leukemia. 2007;109(2):399-404.
92. Fathi AT, Karp JEJCor. New agents in acute myeloid leukemia: beyond cytarabine and anthracyclines. 2009;11(5):346-52.
93. Aggarwal BB, Surh Y-J, Shishodia S. The molecular targets and therapeutic uses of curcumin in health and disease: Springer Science & Business Media; 2007.
94. Kuttan R, Bhanumathy P, Nirmala K, George MJCl. Potential anticancer activity of turmeric (*Curcuma longa*). 1985;29(2):197-202.
95. Cragg GM, Newman DJJJoe. Plants as a source of anti-cancer agents. 2005;100(1-2):72-9.
96. Ferrari C, Torres EJB, Pharmacotherapy. Biochemical pharmacology of functional foods and prevention of chronic diseases of aging. 2003;57(5-6):251-60.
97. Chuang C-C, McIntosh MKJAron. Potential mechanisms by which polyphenol-rich grapes prevent obesity-mediated inflammation and metabolic diseases. 2011;31:155-76.
98. Frederiksen H, Mortensen A, Schröder M, Frandsen H, Bysted A, Knuthsen P, et al. Effects of red grape skin and seed extract supplementation on atherosclerosis in Watanabe heritable hyperlipidemic rabbits. 2007;51(5):564-71.
99. Zunino SJJTJon. Type 2 diabetes and glycemic response to grapes or grape products. 2009;139(9):1794S-800S.
100. Kang JS, Lee WK, Lee CW, Yoon WK, Kim N, Park S-K, et al. Improvement of high-fat diet-induced obesity by a mixture of red grape extract, soy isoflavone and L-carnitine: implications in cardiovascular and non-alcoholic fatty liver diseases. 2011;49(9):2453-8.
101. Tao X, Lipsky PEJRDCoNA. The Chinese anti-inflammatory and immunosuppressive herbal remedy *Tripterygium wilfordii* Hook F. 2000;26(1):29-50.
102. Kaur A, Nain P, Nain JJIJPPS. Herbal plants used in treatment of rheumatoid arthritis: a review. 2012;4(Suppl 4):44-57.
103. Ogle NJAJoMH. Green tea *Camellia sinensis*. 2009;21(2):44.
104. Lansky EP, Newman RAJJoe. *Punica granatum* (pomegranate) and its potential for prevention and treatment of inflammation and cancer. 2007;109(2):177-206.
105. Li Y, Qi Y, Huang TH, Yamahara J, Roufogalis BDJD, Obesity, Metabolism. Pomegranate flower: a unique traditional antidiabetic medicine with dual PPAR- α / γ activator properties. 2008;10(1):10-7.
106. Opara LU, Al-Ani MR, Al-Shuaibi YSJE, Technology B. Physico-chemical properties, vitamin C content, and antimicrobial properties of pomegranate fruit (*Punica granatum* L.). 2009;2(3):315-21.
107. Orhan I, Şenol F, Kartal M, Dvorska M, Žemlička M, Šmejkal K, et al. Cholinesterase inhibitory effects of the extracts and compounds of *Maclura pomifera* (Rafin.) Schneider. 2009;47(8):1747-51.
108. Chowdhury R, Hasan CM, Rashid MAJF. Antimicrobial activity of *Toona ciliata* and *Amoora rohituka*. 2003;74(1):155-8.
109. Li S, Lei Y, Jia Y, Li N, Wink M, Ma YJP. Piperine, a piperidine alkaloid from *Piper nigrum* re-sensitizes P-gp, MRP1 and BCRP dependent multidrug resistant cancer cells. 2011;19(1):83-7.
110. Greco WR, Bravo G, Parsons JCJPr. The search for synergy: a critical review from a response surface perspective. 1995;47(2):331-85.
111. Cai H, Dai F, Min Q, Shi M, Miao J, Luo RJDjydxbaJotfmcOP. Clinical study of the effects of radiotherapy in combination with traditional Chinese medicine on non-small cell lung cancer. 2002;22(12):1112-3.
112. Efferth T, Davey M, Olbrich A, Rücker G, Gebhart E, Davey RJBC, Molecules, et al. Activity of drugs from traditional Chinese medicine toward sensitive and MDR1-or MRP1-overexpressing multidrug-resistant human CCRF-CEM leukemia cells. 2002;28(2):160-8.
113. Janku F, Hong DS, Fu S, Piha-Paul SA, Naing A, Falchook GS, et al. Assessing PIK3CA and PTEN in early-phase trials with PI3K/AKT/mTOR inhibitors. 2014;6(2):377-87.
114. Bozic I, Reiter JG, Allen B, Antal T, Chatterjee K, Shah P, et al. Evolutionary dynamics of cancer in response to targeted combination therapy. 2013;2:e00747.
115. Katzung BJCGS. Basic & Clinical Pharmacology Appleton & Lange Norwalk. 1995.
116. Eid SY, El-Readi MZ, Wink MJP. Synergism of three-drug combinations of sanguinarine and other plant secondary metabolites with digitonin and doxorubicin in multi-drug resistant cancer cells. 2012;19(14):1288-97.
117. Embree L, Gelmon K, Tolcher A, Hudon N, Heggie J, Dedhar C, et al. Pharmacokinetic behavior of vincristine sulfate following administration of vincristine sulfate liposome injection. 1998;41(5):347-52.
118. Embree L, Gelmon KA, Tolcher AW, Hudon NJ, Heggie JR, Dedhar C, et al. Validation of a high-performance liquid chromatographic assay method for quantification of total vincristine sulfate in human plasma following administration of vincristine sulfate liposome injection. 1997;16(4):675-87.
119. Raj TAS, Smith AM, Moore ASJlJon. Vincristine sulfate liposomal injection for acute lymphoblastic leukemia. 2013;8:4361.
120. Shah NN, Merchant MS, Cole DE, Jayaprakash N, Bernstein D, Delbrook C, et al. Vincristine sulfate liposomes injection (VSLI, Marqibo®): results from a phase I study in children, adolescents, and young adults with refractory solid tumors or leukemias. 2016;63(6):997-1005.
121. Schiller GJ, Damon LE, Stock W, Coutre SE, Hsu P, Prasad L, et al. Marqibo®, Vincristine Sulfate Liposome Injection, for the Treatment of Advanced, Relapsed or Refractory Philadelphia Chromosome-Negative (Ph-) Acute Lymphoblastic Leukemia (ALL) in an Adolescent Young Adult (AYA) Population. Am Soc Hematology; 2015.
122. Liu C, Zheng J, Deng L, Ma C, Li J, Li Y, et al. Targeted intracellular controlled drug delivery and tumor therapy through in situ forming Ag nanogates on mesoporous silica nanocontainers. 2015;7(22):11930-8.
123. Tatar A-S, Nagy-Simon T, Tomuleasa C, Boca S, Astilean SJJoCR. Nanomedicine approaches in acute lymphoblastic leukemia. 2016;238:123-38.
124. Bhushan S, Kakkar V, Pal HC, Mondhe D, Kaur IPJC-bi. The augmented anticancer potential of AP9-cd loaded solid lipid nanoparticles in human leukemia Molt-4 cells and experimental tumor. 2016;244:84-93.
125. Muqbil I, Masood A, Sarkar FH, Mohammad RM, Azmi ASJC. Progress in nanotechnology based approaches to enhance the potential of chemopreventive agents. 2011;3(1):428-45.
126. Loureiro JA, Gomes B, Coelho MA, Carmo Pereira Md, Rocha SJN. Targeting nanoparticles across the blood-brain barrier with monoclonal antibodies. 2014;9(5):709-22.

127. Jain KKJN. Nanobiotechnology-based strategies for crossing the blood–brain barrier. 2012;7(8):1225-33.
128. Ornes SJPotNAoS. Antibody–drug conjugates. 2013;110(34):13695-129. Gerber H-P, Koehn FE, Abraham RTJNpr. The antibody-drug conjugate: an enabling modality for natural product-based cancer therapeutics. 2013;30(5):625-39.
130. Langer M, Kratz F, Rothen-Rutishauser B, Wunderli-Allenspach H, Beck-Sickinger AGJJomc. Novel peptide conjugates for tumor-specific chemotherapy. 2001;44(9):1341-8.
131. Ahlert J, Shepard E, Lomovskaya N, Zazopoulos E, Staffa A, Bachmann BO, et al. The calicheamicin gene cluster and its iterative type I enediyne PKS. 2002;297(5584):1173-6.
132. Zaro JL. Mylotarg: revisiting its clinical potential post-withdrawal. Antibody-Drug Conjugates: Springer; 2015. p. 179-90.
133. Norsworthy KJ, Ko CW, Lee JE, Liu J, John CS, Przepiorka D, et al. FDA Approval Summary: Mylotarg for Treatment of Patients with Relapsed or Refractory CD33-Positive Acute Myeloid Leukemia. 2018;23(9).134. Ikemoto N, Kumar RA, Ling T-T, Ellestad GA, Danishefsky SJ, Patel DJJPotNAoS. Calicheamicin-DNA complexes: warhead alignment and saccharide recognition of the minor groove. 1995;92(23):10506-10.
135. Younes A, Gopal AK, Smith SE, Ansell SM, Rosenblatt JD, Savage KJ, et al. Results of a pivotal phase II study of brentuximab vedotin for patients with relapsed or refractory Hodgkin's lymphoma. 2012;30(18):2183.
136. Sievers EL, Senter PDJArom. Antibody-drug conjugates in cancer therapy. 2013;64.
137. Luesch H, Moore RE, Paul VJ, Mooberry SL, Corbett THJJJoNP. Isolation of dolastatin 10 from the marine cyanobacterium *Symploca* species VP642 and total stereochemistry and biological evaluation of its analogue symplostatin 1. 2001;64(7):907-10.
138. Vyas HK, Pal R, Vishwakarma R, Lohiya NK, Talwar GJO. Selective killing of leukemia and lymphoma cells ectopically expressing hCG β by a conjugate of curcumin with an antibody against hCG β subunit. 2009;76(2):101-11.
139. Cragg GM, Newman DJ, Yang SSJJJonp. Natural product extracts of plant and marine origin having antileukemia potential. The NCI experience. 2006;69(3):488-98.