Detection Of 11q23 Gene Rearrangement In Children With Acute Lymphoblastic Leukemia And Its Association With Demographic Data and Response To Initial Chemotherapy On The Seventh Day Of Induction

Alghasi A¹*, Pedram M¹, Saki N¹, Salari F¹, Jaseb K¹, Aminasnafi A¹, Yousefi H¹, Saki Malehi A², Noroozi F¹, Moeinzadeh L¹, Galehdari H³, Malekaskar AM⁴

- 1. Health Research Institute, Research Center of Thalassemia & Hemoglobinopathy, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran
- 2. Department of Biostatistics and Epidemiology, School of Public Health, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran
- 3. Department of Genetics, Shahid Chamran University, Ahvaz, Iran
- 4. Department of Genetics, Qom University of Medical Sciences, Qom, Iran

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Abstract

Background: Acute lymphoblastic leukemia (ALL) is the most common form of childhood cancer leading to cancer-related death in children. Most infants with ALL harbor recurring structural chromosomal rearrangements that are important initiating events in leukemogenesis but are insufficient to explain the biology and heterogeneity of the disease. Mixed-lineage leukemia-rearrangement (MLL-rearrangement) at 11q23 occurs in at least two-thirds of infants with ALL. The most common MLL rearrangements are t(4;11)(q21;q23)/MLL-AFF1 (AF4) found in approximately 50% of patients.

Methods: Forty children with ALL were enrolled in our study. 11q23 rearrangement and its association with other prognostic factors such as age, sex, initial WBC, organomegaly, immunophenotype, and therapeutic response on the seventh day of induction were studied.

Results: Four patients including three (11.5%) boys and one (7.1%) girl were positive for 11q23 translocation. There was no association between 11q23 rearrangement and sex, age, and initial WBC counts. None of the patients with 11q23 translocation showed blast count less than 5% in the bone marrow on the seventh day of induction (P=0.002). **Conclusion:** There was a significant correlation between 11q23 translocation with lack of initial response to chemotherapy.

Keywords: Acute lymphoblastic leukemia, 11q23 translocation, Cytogenetic, Infant acute lymphoblastic leukemia, prognosis, Induction failure

Introduction

Acute lymphoblastic leukemias (ALL) is the most common type of childhood cancer representing 80–85% of all leukemias ¹. ALL in infants often presents with a number of unfavorable clinical features such as hyperleukocytosis, organomegaly, and central nervous system (CNS) involvement ².

Cytogenetic abnormalities have been described in 80% of childhood ALLs ^{3,4}.

Some chromosomal abnormalities are well known independent prognostic factors in childhood ALL and are used to predict the outcome⁵. Favorable biological characteristics (TEL-AML1 fusion,

^{*}Corresponding Author: Alghasi A, Email: arashalgasi@yahoo.com

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hyperdiploidy) are seldom found in infant ALL whereas myeloid-associated antigen co-expression is a common finding ^{6,7}. Infantile ALL is commonly associated with MLL gene rearrangements and pro-B (pre-preB/immature/CD10-negative precursor-B) immunophenotype ⁸.

MLL gene rearrangements arise from fusions of this gene at 11g23 with a large number of partner genes (table 1). In ALL, the most common gene partner is the AF4 gene on chromosome 4q21, resulting in a t(4; 11) (q21; q23) rearrangement^{9,10}. It is not clear which of the features associated with infant ALL specifically contribute to the poor therapeutic response 11. Children with MLL gene abnormalities, and more specifically MLL-AF4 fusion, seem to have a poor prognosis 11,12. Early in vivo response to treatment (such as prednisolone) was found to be predict the outcome in infant ALL in the German Berlin-Frankfurt-Munster (BFM) studies 13. However, it is unclear which of the factors (age, MLL rearrangements, or immunophenotype) is most important. The common 11q23 translocation and its partner genes are listed in table 1.

In this study, we aimed to investigate 11q23 rearrangements and their association with other factors such as age, sex, initial WBC, organomegaly, immunophenotype, and therapeutic response. We particularly aimed to assess whether MLL aberrations could be a reliable prognostic factor in childhood ALL.

Patients and Methods

Forty patients with ALL referred to Ahvaz Shafa Hospital during 2012-2013 were enrolled in this study. The study population consisted of a group of children with pre-B ALL that had been diagnosed by morphological and flowcytometric assays and were treated with a similar therapeutic protocol in our institute. In addition, bone marrow samples were obtained on the seventh day of the induction to assess residual blasts. Bone marrow samples were also studied for karyotype and cytogenetic abnormalities.

All procedures involving human participants were in accordance with the ethical standards of the local ethics committee of Ahvaz Jundishapur University of Medical Sciences (REC.1392.209) and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. Informed consent was obtained from all individual

participants included in the study.

Cytogenetic analysis

Preparation of chromosomes was done by bone marrow culture on the slide, i.e. the karyotyping technique. The obtained chromosomes were stained by GIMSA and later denaturalized by trypsin. Then mitosis analysis was done for detecting numerical and structural chromosome aberrations.

Fluorescence in situ hybridization (FISH) analyses

FISH analyses were performed using a locus-specific identifier (LSI) MLL dual-color DNA probe which hybridizes to the MLL gene with a Spectrum Green-labeled probe on the centromeric side (5') and a spectrum orange-labeled probe to the telomeric side (3') of the MLL gene breakpoint region. These two signals fuse to one yellow signal in interphase nuclei of normal cells. In cases with MLL rearrangement one yellow signal is present on the normal chromosome 11 and one green and one orange signal are detected on the translocation partner chromosomes.

Descriptive data analysis was conducted using SPSS software. Chi-square test was used to assess the correlation between variables , and P<0.05 was considered significant.

Results

Forty infants with ALL were included in this study. All patients immunophenotypically were pre-B ALL. The median age of the patients was 48 months (range: 11-48 months); 26 (65%) of the patients were boys and 14 (35%) were girls. Initial white blood cell (WBC) counts of the patients ranged from 0.1-230×10⁹/L (median=23.66×10⁹/L). of the 40 studied infants, 12 (30%), two (5%) and four (10%) had splenomegaly, hepatomegaly and hepatosplenomegaly, respectively. FISH analysis showed that 23 (88.5%) boys and 13 (92.9%) girls had no chromosomal abnormalities. Four patients including three (11.5%) boys and one (7.1%) girl were positive for 11q23 translocation. There was no gender difference for 11q23 translocation among these four cases (P=0.6). Likewise, there was no significant difference among patients positive for 11q23 translocation with respect to age (P=0.18).

Considering BFM classification, 23 and 17 patients were diagnosed with L1 and L2 morphology, respectively. Of the four patients who had 11q23

Table 1: Most frequent translocations of the MLL (11q23) gene in patients with ALL

| Translocations | Breakpoint regions | Frequency in ALL patients | MLL fusion partners | Function of partner gene | Outcome | Reference | |
|--|---|---------------------------------------|---|--|---|-------------|--|
| t(1;11) | (p32;q23) (q21;q23) | Rare | AF-1p AF-1q | Putative signal transduction Growth factor | Poor | (14) | |
| t(4;11) | (q21;q23) | Infant: 50- 70% Adult : 3-7% | A-F4 (AFF1) FLJ-10849 | Transcription activator | poor (Age<1: worse outcome) | (4, 5, 15) | |
| t(6;11) | (q21;q23) (q27;q23) | Rare | AF-6q21 AF-6 | Transcription factor Signal transduction | Poor | (14) | |
| t(9;11) | (p22;q23) (p34;q23) (p34;q23) | Rare | <i>MLLT-</i> 3(<i>AF-9</i>) AF-9q34 FBP-17 | Transcription activator Negative regulator of RAS Telomere maintenance | Poor | (4, 14, 16) | |
| (p11.2;q23) t(10;11) (p12;q23) (q21;q23) | | Rare | ABI-1 AF-10 LCX | Cell growth inhibitor Transcription activator Methyltransferase domain | Poor | (4, 14) | |
| t(11;19) | (q23;p13.1) (q23;p13.3) (q23;p13.3) | 13% | ELL EEN ENL | Regulation of cell growth and survival Signal transduction Transcription activator | poor (Age<1: worse outcome Age< 6 months: inferior treatment outcome) | (14, 16) | |
| (q23;p13) t(11;17) (q23;q21) (q23;q25) | | Rare | GAS7 AF17 MSF | Growth-arrest specific protein Transcription factor cycle regulation / Signal transduction | Poor | (14, 16) | |

MLL: Mixed-lineage leukemia; AF-1: Associated factor-1; FBP-17: Formin-bindenden Proteins 17; ABI-1: Abelson interacting protein 1; LCX: Leukemia-associated Protein with a CXXC Domain; ELL: eleven-nineteen lysine-rich leukemia protein; EEN: extra eleven nineteen gene; ENL: Eleven Nineteen Leukemia gene; GAS-7: growth arrest-specific-7; MSF: MLL septin-like fusion.

Table 2: Demographic, clinical and laboratory characteristics of patients with ALL

| | Age (month) | Sex | Leukocyte count (*10 ⁹ /L) | Organomegaly | Morphology | Karyotype | FISH | ВМА | CNS |
|----|----------------|-----|---|--------------|------------|------------------------|------|-------|-----|
| 1 | 51.0 | М | 27.2 | NO | L2 | 46xy | N | <5% | - 1 |
| 2 | 48.0 | M | 39.9 | NO | L1 | 46xy | N | <5% | - 1 |
| 3 | 36.0 | M | 49.3 | SM | L1 | 46xy, t(9,11),t(14,22) | N | <5% | - 1 |
| 4 | 36.0 | M | 40.7 | NO | L1 | 46xy | N | <5% | - 1 |
| 5 | 30.0 | M | 12.7 | NO | L1 | 46xy | Р | 5-25% | - 1 |
| 6 | 15.0 | F | 74.9 | NO | L2 | 46xx, t(4,11) | Р | 5-25% | - 1 |
| 7 | 96.0 | M | 7.4 | NO | L1 | 46xy | N | <5% | - 1 |
| 8 | 18.0 | F | 15.4 | SM+HM | L1 | 46xx | Ν | <5% | Ш |
| 9 | 11.0 | M | 18.3 | SM | L1 | 46xy | Ν | <5% | - 1 |
| 10 | 124.0 | M | 8.7 | NO | L2 | 46xy | Ν | 5-25% | - 1 |
| 11 | 80.0 | F | 2.4 | NO | L2 | 46xx | N | 5-25% | - 1 |
| 12 | 73.0 | F | 3.2 | SM | L1 | 46xx | N | 5-25% | - 1 |
| 13 | 24.0 | M | 87.5 | HM | L2 | 46xy,del2 | N | <5% | - 1 |
| 14 | 36.0 | F | 4.8 | NO | L1 | 46xx,t(11;19),del5p | N | <5% | - 1 |
| 15 | 83.0 | M | 3.0 | NO | L2 | 46xy,t(1;11)del15 | N | <5% | - 1 |
| 16 | 52.0 | F | 10.0 | NO | L2 | 46xx | N | <5% | - 1 |
| 17 | 56.0 | M | 2.6 | SM | L1 | 46xy,t(11;19),del18 | N | <5% | - 1 |
| 18 | 40.0 | M | 4.1 | NO | L1 | 46xy,t(4,3) | N | <5% | 1 |
| 19 | 124.0 | M | 1.0 | NO | L2 | 46xy | N | 5-25% | - 1 |
| 20 | 36.0 | M | 7.7 | NO | L2 | 46xy | N | <5% | 1 |
| 21 | 134.0 | F | 3.2 | NO | L1 | 46xx | N | <5% | 1 |
| 22 | 112.0 | M | 12.2 | SM | L2 | 46xy | Р | 5-25% | - 1 |
| 23 | 132.0 | F | 1.2 | NO | L1 | 46xx | N | <5% | - 1 |
| 24 | 28.0 | F | 9.9 | SM | L2 | 46xx | N | 5-25% | - 1 |
| 25 | 12.0 | M | 122.2 | SM+HM | L1 | 46xy | N | <5% | - 1 |
| 26 | 148.0 | M | 2.6 | SM | L1 | 46xy | N | <5% | - 1 |
| 27 | 57.0 | F | 11.1 | SM | L1 | 46xx | N | <5% | - 1 |
| 28 | 60.0 | F | 11.5 | SM | L1 | 46xx | N | <5% | - 1 |
| 29 | 52.0 | M | 23.3 | SM+HM | L1 | 46xy | N | <5% | - 1 |
| 30 | 12.0 | M | 4.6 | SM | | 46xy | Р | >25% | - 1 |
| 31 | 26.0 | M | 1.9 | NO | L2 | 46xy,t(1,14)(p32-34) | N | <5% | - 1 |
| 32 | 22.0 | F | 5.3 | SM | L1 | 46xx,inv16(p13;q22) | N | 5-25% | 1 |
| 33 | 24.0 | M | 32.0 | SM+HM | L2 | 46xy | N | <5% | 1 |
| 34 | 48.0 | M | 14.8 | SM | L2 | 46xy,t(1,11) | N | <5% | - 1 |
| 35 | 148.0 | М | 4.5 | NO | L1 | 46xy,t(1,11)t(4.11) | N | 5-25% | 1 |
| 36 | 24.0 | F | 15.5 | HM | L2 | 46xx,t(2,8),t(4,11) | N | <5% | 1 |
| 37 | 40.0 | М | 0.1 | NO | L1 | 46xy | N | <5% | Í |
| 38 | 36.0 | F | 4.4 | NO | L2 | 46xx | N | 5-25% | Ĺ |
| 39 | 72.0 | М | 230.0 | NO | L1 | 46xx | N | 5-25% | II |
| 40 | 72.0 | M | 15.3 | NO | L2 L1 | 46xy | N | <5% | Ï |

M: Male; F: Female; NO: No organomegaly; SM: Splenomegaly; HM: Hepatomegaly; BM: bone marrow; CNS: central nervous system

rearrangements, L1 morphology was seen in one and L2 in three cases.

The mean initial WBC counts in patients without 11q23 translocation and those positive for MLL gene rearrangement was $9.3\times10^9/L$ (range: $0.1-230\times10^9/L$) and $12.4\times10^9/L$ (range: $4600-74900\times10^9/L$), respectively. There was no significant relationship between WBC count and 11q23 translocation .

By assessing the initial response to chemotherapy based on bone marrow samples obtained on the seventh day of induction, 27 (67.5%) infants had less than 5% blasts, 12 (30%) 5- 25% blasts and one had more than 25% blasts. However, all four patients with 11q23 translocation had more than 5% blasts. There was a positive association between 11q23 translocation and lack of initial response to chemotherapy (P=0.002).

Discussion

There was a significant correlation between 11q23 translocation and lack of initial response to chemotherapy in our study. However, the associations with other variables such as sex, age, and WBC counts were not meaningful. These insignificant results could have also been due to the small sample size of our study.

11q23 translocation is a well known poor prognostic factor which is observed in 2-4% of childhood ALL but is expressed in 80% of infants with ALL¹⁷. Most children who harbor 11q23 translocations show (4; 11) (q21; q23) abnormality¹⁸.

In a cytogenetic study of cancer cells in 56 children, 21 children indicated t(4,11). Out of 16 patients with a normal karyotype, two cases showed t(1,19) (q32; p13), one case had more than 50 chromosomes and 9 patients had non-recurring structural abnormalities 18. In a cytogenetic study on 39 infant ALL, 12 (31%) patients had translocation t(4,11)(g21;g23) and EFS was remarkably reduced in these patients. Furthermore, five cases had chromosomal-breakage in 11q23 that this group had longer survival than the previous group. In total, structural disorders were seen in 27 out of 28 patients with abnormal karyotype 19. In another study, 30 infants suffering from lymphoblastic leukemia were investigated, of which 14 cases had cytogenetic abnormality in 11q23. The results of this study showed that molecular rearrangement in 11q23 specifically has a relationship with adverse prognostic factors such as age less than six months,

hyperleukocytosis, CD10-phenotype, and an initial treatment failure ²⁰.

In a large study on 6238 patients with B-precursor ALL, four risk groups were considered and a five-year survival rate was determined and unfavorable prognostic factors were assessed. 11q23 rearrangement was an adverse prognostic factor that put the patients in the high-risk group and was associated with reduced five-year survival rate ²¹.

In another study on 37 infants with ALL who were less than 18 months old, the effect of 11q23 on prognosis was studied and similar results with previous studies were observed. A total of 18 patients had ALL. Analysis of the results revealed 67% were girls; 50% of them had hyperleukocytosis and 39% had a rearrangement in 11q23. 5-year survival rate of these patients was 14±12%. Nineteen patients had myeloid leukemia and 53% of the cases had rearrangement in 11q23. This group had a very poor prognosis and five-year survival rate was reported to be 20±9% ²².

Raimondi and colleagues studied 785 cases in whom 17 had structural abnormality in the 11q23; 14 patients were del(11)(q23) and three patients showed inv(11)(p12q23). Unlike previous studies, these patients had a better prognosis and fewer leukocytes. This study showed that deletion or reversal in chromosome 11q23 has a better prognosis than patients with 11q23 translocation²³.

In our study, age of the patients was in a higher range than the previous studies. Also on the seventh day of induction all children with 11q23 abnormality had a blast count higher than 5% that represents a poor prognostic factor. Also, it was observed that among patients with 11q23, t(4,11) was among the most common partners of 11q23.

In summary, MLL rearrangements are associated with poor outcome in pediatric ALL. Here we showed an association between 11q23 rearrangement and lack of initial response to chemotherapy, but no relationship was seen between 11q23 translocation with age and sex. Additional molecular and cytogenetic studies on children with ALL in our country considering the ethnicity of Iranian children are needed to precisely find the frequency of MLL gene rearrangements and their correlation with other prognostic factors.

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Conflict of interest disclosure

The authors declare no competing financial interests.

References

- Caughey RW, Michels KB. Birth weight and childhood leukemia: A meta-analysis and review of the current evidence. International Journal of Cancer. 2009;124(11):2658-70.
- Pui C-H, Behm FG, Downing JR, Hancock ML, Shurtleff SA, Ribeiro RC, et al. 11q23/MLL rearrangement confers a poor prognosis in infants with acute lymphoblastic leukemia. Journal of Clinical Oncology. 1994;12(5):909-15.
- Martinez-Climent JA, Espinosa III R, Thirman MJ, Le Beau MM, Rowley JD. Abnormalities of chromosome band 11q23 and the MLL gene in pediatric myelomonocytic and monoblastic leukemias: identification of the t (9; 11) as an indicator of long survival. Journal of pediatric hematology/oncology. 1995;17(4):277-83.
- Mullighan CG. Molecular genetics of B-precursor acute lymphoblastic leukemia. The Journal of clinical investigation. 2012;122(10):3407.
- 5. S. Mrozek K, Heerema NA, Bloomfield CD. Cytogenetics in acute leukemia. Blood reviews. 2004;18(2):115-36.
- Harrison CJ. The detection and significance of chromosomal abnormalities in childhood acute lymphoblastic leukaemia. Blood reviews. 2001;15(1):49-59.
- Zemanova Z, Michalova K, Sindelarova L, Smisek P, Brezinova J, Ransdorfova S, et al. Prognostic value of structural chromosomal rearrangements and small cell clones with high hyperdiploidy in children with acute lymphoblastic leukemia. Leukemia research. 2005;29(3):273-81.
- Reaman GH, Sposto R, Sensel MG, Lange BJ, Feusner JH, Heerema NA, et al. Treatment outcome and prognostic factors for infants with acute lymphoblastic leukemia treated on two consecutive trials of the Children's Cancer Group. Journal of Clinical Oncology. 1999;17(2):445-.
- 9. Dobbins SE, Sherborne AL, Ma YP, Bardini M, Biondi A, Cazzaniga G, et al. The silent mutational landscape

- of infant MLL-AF4 pro-B acute lymphoblastic leukemia. Genes, Chromosomes and Cancer. 2013;52(10):954-60.
- Yokoyama A, Lin M, Naresh A, Kitabayashi I, Cleary ML. A higher-order complex containing AF4 and ENL family proteins with P-TEFb facilitates oncogenic and physiologic MLL-dependent transcription. Cancer cell. 2010;17(2):198-212.
- 11. Kumar AR, Yao Q, Li Q, Sam TA, Kersey JH. t (4; 11) leukemias display addiction to MLL-AF4 but not to AF4-MLL. Leukemia research. 2011;35(3):305-9.
- 12. Drake AS, Brady MT, Wang XH, Sait SJ, Earp JC, Ghoshal S, et al. Targeting 11q23 positive acute leukemia cells with high molecular weightmelanoma associated antigen-specific monoclonal antibodies. Cancer immunology, immunotherapy. 2009;58(3):415-27.
- Dördelmann M, Reiter A, Borkhardt A, Ludwig W-D, Götz N, Viehmann S, et al. Prednisone response is the strongest predictor of treatment outcome in infant acute lymphoblastic leukemia. Blood. 1999;94(4):1209-17.
- 14. De Braekeleer M, Morel F, LE BRIS M-J, HERRY A, DOUET-GUILBERT N. The MLL gene and translocations involving chromosomal band 11q23 in acute leukemia. Anticancer research. 2005;25(3B):1931-44.
- 15. Ivanov IC, Jitaru D, Grigore GE, Zlei M, Ivanov AV, Dumitraş S, et al. Infant acute leukemia with lineage switch at relapse expressing a novel t (4; 11)(q21; q23) MLL-AF4 fusion transcript. Revista Română de Medicină de Laborator Vol. 2013;21(1/4).
- Pui C, Chessells J, Camitta B, Baruchel A, Biondi A, Boyett J, et al. Clinical heterogeneity in childhood acute lymphoblastic leukemia with 11q23 rearrangements. Leukemia. 2003;17(4):700-6.
- Karkucak M, Gorukmez O, Yakut T, Baytan B, Gorukmez O, Gunes AM. Molecular Cytogenetic Findings in Cases with Childhood Acute Lymphoblastic Leukemia. International Journal of Hematology & Oncology/UHOD: Uluslararasi Hematoloji Onkoloji Dergisi. 2012;22(2).
- Schad CR, Kraker WJ, Jalal SM, Tallman MS, Londer HN, Cook LP, et al. Use of fluorescent in situ hybridization for marker chromosome identification in congenital and neoplastic disorders. American journal of clinical pathology. 1991 Aug;96(2):203-10. PubMed PMID: 1862775. Epub 1991/08/01. eng.
- 19. Heerema NA, Arthur DC, Sather H, Albo V, Feusner J, Lange BJ, et al. Cytogenetic features of infants

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- less than 12 months of age at diagnosis of acute lymphoblastic leukemia: impact of the 11q23 breakpoint on outcome: a report of the Childrens Cancer Group. Blood. 1994;83(8):2274-84.
- Chen C-S, Sorensen P, Domer P, Reaman G, Korsmeyer S, Heerema N, et al. Molecular rearrangements on chromosome 11q23 predominate in infant acute lymphoblastic leukemia are associated with specific biologic variables and poor outcome. Blood. 1993;81(9):2386-93.
- 21. Schultz KR, Pullen DJ, Sather HN, Shuster JJ, Devidas M, Borowitz MJ, et al. Risk-and response-based classification of childhood B-precursor acute lymphoblastic leukemia: a combined analysis of prognostic markers from the Pediatric Oncology Group (POG) and Children's Cancer Group (CCG). Blood. 2007;109(3):926-35.
- 22. Cabrera M, Campbell M, Quintana J, Undurraga M, Ford A, Greaves M. [Clinical significance and frequency of the 11q23/MLL genetic molecular alteration in Chilean infants with acute leukemia]. Revista medica de Chile. 2001;129(6):634-42.
- 23. Raimondi S, Frestedt J, Pui C, Downing J, Head D, Kersey J, et al. Acute lymphoblastic leukemias with deletion of 11q23 or a novel inversion (11)(p13q23) lack MLL gene rearrangements and have favorable clinical features. Blood. 1995;86(5):1881-6.